

Toxicity of south Morocco *Rosmarinus officinalis* essential oil: antibacterial and histopathological effects

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(Reçu le 22/10/2004 ; Accepté le 16/12/2004)

دراسة الخاصية السمية للزيت الأساسي للأزير المغربي : فعاليتها كمضادات البكتيريا ومضادات الأكسدة

الأزير شجيرة عطرية تمتلك خاصيات عديدة وهي معروفة بفعاليتها كمضادات الميكروبات ومضادات الأكسدة. الهدف من هذا البحث هو دراسة الخاصيات السمية للزيت الأساسي للأزير وذلك بواسطة تجربتين مختلفتين. تم تحليل التركيبة الكيميائية لهذا الزيت بواسطة الإستشراب في الطور الغازي و دراسة نشاط مضاد البكتيريا بتقنية المكرو أطموسفير. تم البحث عن الإصابات النسيجية بواسطة التشريح المرضي النسيجي لأعضاء الفئران (عدها 20) بعد إطعامها بواسطة 50 ميكرو لتر في الكيلو غرام من الأتلة الجسمية للحيوان، كقياس يومي، لمدة سبعة أيام مقارنة مع فئران تم تغذيتها بالماء المقطر بنفس الكمية وخلال نفس المدة. أظهرت النتائج أن المكونات الغالبة للزيت الأساسي للأزير هي سينول (42%)، البنين (11,92%) و الكونفر (13,99%) تتغير الكمية الدنيا للتثبيط عند البكتيريا المجربة من 50 إلى 90 ميكرو لتر. أظهرت الدراسة التشريحية المرضية للأنسجة توسع السنخ الرئوي وتضخم خلايا الغدة الكظرية.

الكلمات المفتاحية : الأزير- الزيت الأساسي - نشاط مضاد البكتيريا - التشريح النسيجي - التسمم

Toxicité de l'huile essentielle de *Rosmarinus officinalis* du sud du Maroc: effets antibactérien et histopathologique

Le romarin (*Rosmarinus officinalis*) possède de nombreuses propriétés médicinales. Il est connu pour ses effets antimicrobiens et anti-oxydatifs. Le but de ce travail est d'étudier la toxicité de son huile essentielle (HE) via un test antibactérien selon la technique de micro-atmosphère et un autre histopathologique au niveau des organes de souris (*Swiss albinos*). L'analyse de la composition chimique de cette HE a été effectuée par CPG. Les constituants majoritaires de l'HE de *R. officinalis* sont le 1,8-cinéole (42%), l'alpha-pinène (11,92%) et le camphre (13,99%). Les quantités minimales inhibitrices des souches bactériennes testées varient entre 40 µl et 90 µl. Au plan histopathologique, une dilatation des alvéoles pulmonaires et une hypertrophie des cellules corticales et médullaires des surrénales ont été relevées. En conclusion, l'HE de *R. officinalis* ne présente pas de toxicité à la dose 50 µl/g mais possède des effets stimulants des glandes surrénales et du système respiratoire.

Mots clés: Cytotoxicité - Huile essentielle - *Rosmarinus officinalis* - Activité antibactérienne - Histopathologie - Errachidia - Maroc

Toxicity of south Morocco *Rosmarinus officinalis* essential oil: antibacterial and histopathological effects

Rosemary is an aromatic plant that have several medicinal properties, which is essentially used for its antimicrobial and anti-oxidant effects. The goal of this work is to study the toxicity of south Morocco *Rosmarinus officinalis* essential oil by antibacterial by micro-atmospheric technique and histopathologic tests on various organs of mice (*Swiss albinos*). Essential oil was analyzed by gas chromatography (GC). The main components detected in our essential oil were 1,8-cineole (42%), alpha-pinene (11.92%) and camphre (13.99%). Minimal inhibitory quantities of *R. officinalis* essential oil are ranged from 40 µl to 90 µl. Histopathological study showed a pulmonary alveolar dilation and hypertrophy of cortical and medullary suprarenal gland cells. In conclusion, *R. officinalis* essential oil do not present a serious toxicity at 50 µl/g dose but induce stimulating effects on respiratory system and suprarenal glands.

Key words: Cytotoxicity - Essential oil - *Rosmarinus officinalis* - Antibacterial activity - Histopathology - Errachidia - Morocco

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INTRODUCTION

Rosemary (*Rosmarinus officinalis*), an evergreen shrub, is one of the herb spices of the labiatae family cultivate among the wild world.

It have been used in cosmetics and in folk medicine as an antispasmodic of renal colic, for relieving respiratory disorders or to stimulate hair growth (Lemonica *et al.*, 1996; Al-Sereiti *et al.*, 1999; Munne-Bosch *et al.*, 2001). It have a potential therapeutic to prevention and treatment of bronchial asthma, inflammatory diseases, atherosclerosis and cancer (Offord *et al.*, 1995; Hui-Hui *et al.*, 2001; Sotelo-felix *et al.*, 2002; Mimica-Durck *et al.*, 2003).

The rosemary anti-oxidant properties is attributed to its diterpenoids, flavonoids, triterpenoids and phenolic acids constituents (Calabrese *et al.*, 2001; Munne *et al.*, 2001; Kim *et al.*, 2003; Ponce *et al.*, 2004). Volatile compound essential oil of secondary metabolism plant may act as phytoprotective agents defending some species of conifer from herbivore and pathogen attack (Gijzen *et al.*, 1991; Faleiro *et al.*, 1999).

Their bioactive components have recently gained momentum in many pharmaceutical and food processing applications (Cowan., 1999; Silva *et al.*, 2003; Vadar-Unlu *et al.*, 2003). They have insecticide, antifungal and antibacterial activities (Pattanaik *et al.*, 1996; Dorman *et al.*, 2000; Benkeblia, 2004).

Several studies have proved that rosemary essential oil possess anti-oxidative and antimicrobial activities which are using for food preservation and human microbial diseases control (Lis-Balchin, 1997; Mondello *et al.*, 2003).

The aim of this work was to determine the composition of Moroccan *Rosmarinus officinalis* essential oil and to study its toxicity by antibacterial and histopathological tests.

MATERIAL & METHODS

1. Plant material

Rosemary (*Rosmarinus officinalis*) samples were collected from Errachidia, Morocco in May 2002. The specimen identification was realized in Biochemistrical Laboratory of the "Institut Agronomique et Vétérinaire Hassan II".

2. Essential oil extraction

Rosemary was submitted for 3 hours to hydrodistillation using a Clevenger-type apparatus. The essential oil was dissolved in n-hexane (10% v/v) before gas chromatography (GC) analysis.

3. Gas chromatography Analysis

The essential oil was analyzed by capillary gas chromatography (Chromapack Cp 9001) equipped with a SE54DF (30 m x 0.25 mm) capillary column. The column temperature was programmed initially at +50°C (isothermal for 5 min), then gradually increased to +230°C (isothermal for 10 min) at +4°C/min rate. A flame ionization detector (FID) was used for routine quantitative analysis. Detector and injector temperature were at respectively +235°C and +240°C. The nitrogen carried gas was adjusted at a flow rate of 1 ml/min.

4. Antibacterial assay: micro-atmosphere method

The micro-organisms used were *Escherichia coli* CIP54127, *Proteus vulgaris* CIP5860T and *Klebsiella pneumoniae* CIP8291T which were a gift from a Pasteur Institute. We have also used *Escherichia coli*, *Proteus vulgaris* and *Salmonella enteritidis* strains which were isolated from patients in Pasteur Institute biological center. These bacteria were selected because they are frequently reported in human infection and are multiresistant to several antibiotics. Strains were maintained in Kligler agar at +4°C.

Bacteria inoculate were prepared by growing cells in Tryptic Soy Broth for 24 h at +37°C. The cell suspensions were diluted with peptone water to provide initial cell counts of about 10^7 to 10^8 colony forming unit (CFU)/ml.

A suspension of the tested micro-organism (5 µl at 10^8 cells/ml) was distributed on the C.E.L.D. agar (Cystine-Lactose-Electrolyte-Déficient) surface. Filter paper discs (20 mm diameter) were impregnated with various quantities of the essential oils (from 0 µl to 100 µl with 5 µl of increment), placed in the cover of the Petri box then incubated at +37°C for 24h. All the tests were performed induplicate and repeated triplicate.

5. Histopathological study

5.1. Animals

Swiss albino mice, six weeks old, purchased from Sciences Faculty, University Hassan II Ain Chock Casablanca, were housed in plastic cages in a conditioned air room ($+22 \pm 2^\circ\text{C}$, humidity $55 \pm 10\%$) and given food and water freely.

5.2. Gavage

Experimental group of mice (10 males and 10 females) were fed with $50 \mu\text{l/g}$ of *Rosmarinus officinalis* essential oil during 7 days and the control group (5 males and 5 females), received the buffer water during the same period.

5.3. Secondary effects

The observation of the general state and the mice mortality has been followed during the 7 days of treatment. Every day, all animals have been weighted and their water and food consumption has been evaluated.

5.4. Microscopic analysis

The mice were sacrificed and the following organs: liver, kidney, brain, spleen, lung, bowel, stomach, heart, testicular, suprarenal glands were removed, fixed in Bouin and embedded in paraffin. $4\text{-}5 \mu\text{m}$ sections were stained with hematein-eosin, then examined under light microscopy (Olympus-BH-2).

RESULTS

1. Chemical composition of essential oil

Hydrodistillation of *Rosmarinus officinalis* dried plant yielded 0.85% (volume/weight) of essential oil (calculated per weight of dried material). GC analysis of the crude oil resulted in the identification of seventeen components representing 95.75% of the total components of essential oil from *Rosmarinus officinalis* (Table 1).

2. Antibacterial activity

Results of antibacterial activity of *Rosmarinus officinalis* essential oil against bacteria is presented in table 2. Minimal inhibitory quantities are ranged from $40 \mu\text{l}$ to $90 \mu\text{l}$ for all strains.

Table 1. Main components (%) of *Rosmarinus officinalis* essential oil. Compounds listed in order of elution. R_t : retention time (as minutes)

	Components	Rt	Percentage
1	α -pinene	7.55	11.92
2	camphene	8.09	4.55
3	β -pinene	9.22	7.71
4	β -myrcene	9.94	1.41
5	p-cymene	11.12	0.93
6	1,8-cineole	11.49	42.00
7	γ -terpinene	12.66	0.86
8	terpinolene	13.85	0.39
9	linalol	14.39	0.88
10	camphor	15.98	13.99
11	borneol	16.81	3.57
12	1-terpinen-4-ol	17.35	0.81
13	α -terpineol	17.86	2.40
14	berny-acetate	21.44	0.73
15	β -caryophyllen	26.07	3.60

Table 2. Minimal inhibitory quantities of *Rosmarinus officinalis* essential oil obtained using micro-atmospheric technique

Strains	Minimal inhibitory quantities (μl)
<i>Escherichia coli</i> CIP54127	65
<i>Proteus vulgaris</i> CIP5860T	40
<i>klebsiella pneumoniae</i> CIP8291T	90
<i>Proteus vulgaris</i>	40
<i>Salmonella enteritidis</i>	60
<i>Escherichia coli</i>	70

3. Histopathological study

Mice treated by *Rosmarinus officinalis* essential oil showed scratching of the muzzle and bewilderment. These signs disappear three hours after the gavage. In the fifth day of treatment, we noted a somnolence and a reduction of the locomotors and the escarping activity.

However, mice do not present any respiratory difficulty during the treatment and no mortality has been observed during all the study.

Weight evaluation and consumption of food and water did not present a very noticed variations (Figures 1, 2 & 3).

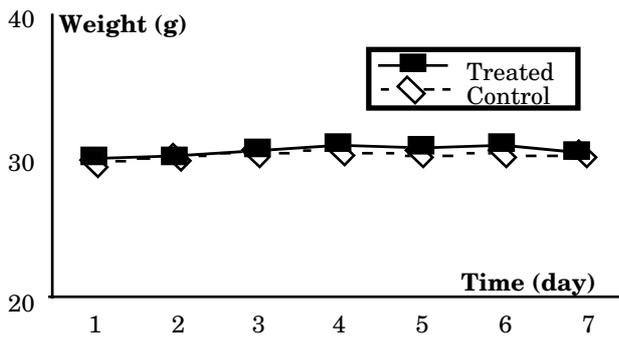


Figure 1. Weight variation of the Swiss albinos mice (n=30) during daily treatment (7 days) with 50 µl/g body weight of *Rosmarinus officinalis* essential oil

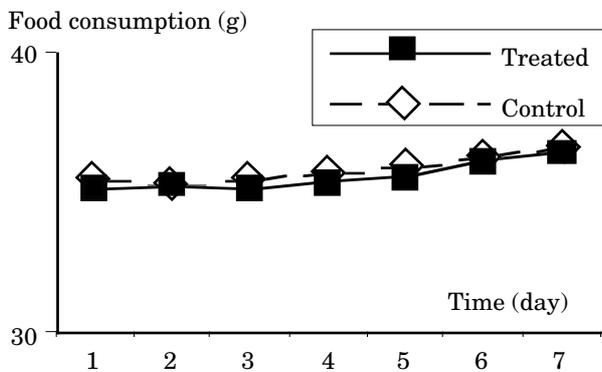


Figure 2. Variation of food consumption of the Swiss albinos mice (n=30) during daily treatment (7 days) with 50 µl/g body weight of *Rosmarinus officinalis* essential oil

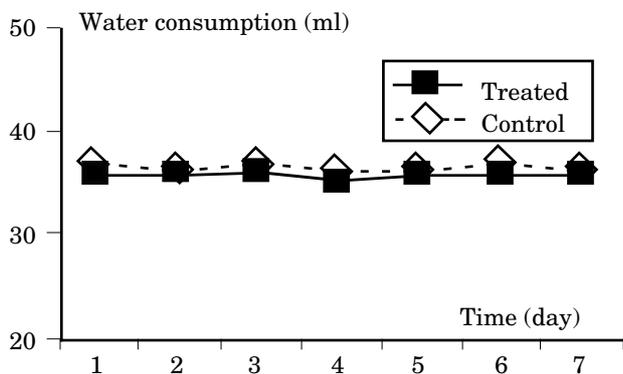


Figure 3. Variation of water consumption of the Swiss albinos mice (n=30) during daily treatment (7 days) with 50 µl/g body weight of *Rosmarinus officinalis* essential oil

The macroscopic analysis of the removed organs during the dissection revealed that the treated lungs were more red then the controls. All the other organs have a normal aspect.

The microscopic analysis of the treated organs showed clearly a pulmonary alveolar dilation (Figure 4) and a cortical and a medullary suprarenal cells hypertrophy (Figure 5).

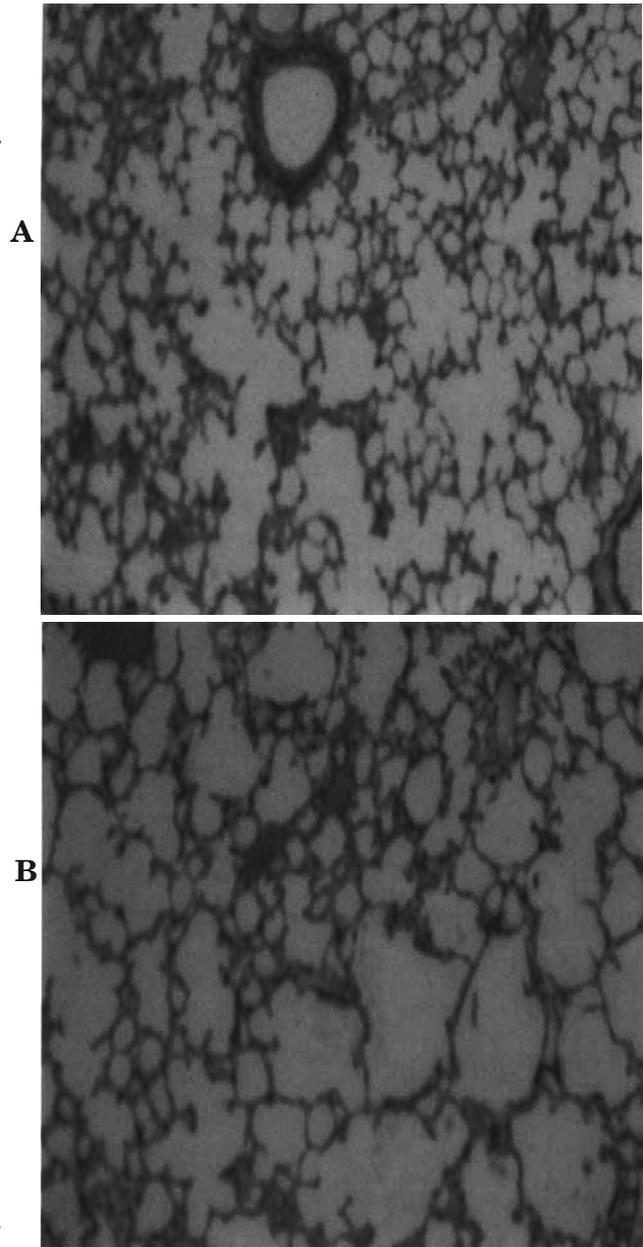


Figure 4. Photomicrographs (Gx40) of the Swiss albinos lung (n=30). Section from control (A) and mice treated with 50 µl/g body weight of *Rosmarinus officinalis* essential oil (B), were stained with hematein-eosine

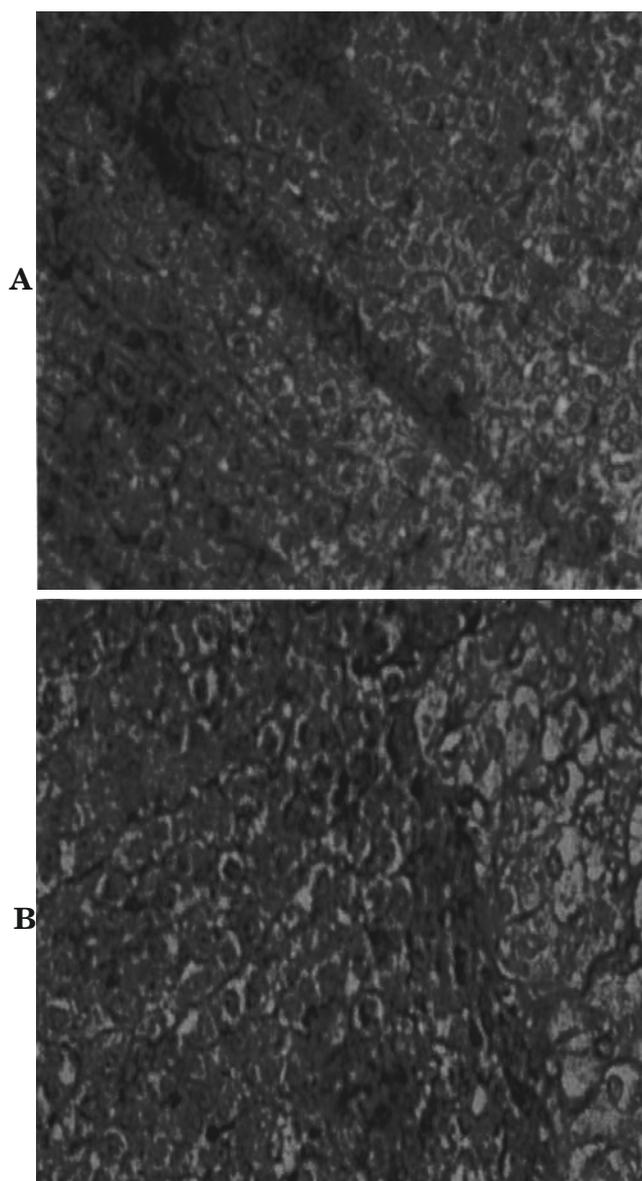


Figure 5. Photomicrographs (Gx40) of the Swiss albinos suprarenal glands (n=30). Section from control (C) and mice treated with 50 µl/g body weight of *Rosmarinus officinalis* essential oil (D), were stained with hematein-eosine

The results of histopathologic analysis of the other organs revealed no considerable abnormality.

DISCUSSION

Chemical composition of Moroccan *Rosmarinus officinalis* essential oil is in accordance with some published data (Chalchat *et al.*, 1993; Fechtal *et al.*, 2000). The 1,8 cineole (42%), the α -pinene (11.92%) and the camphre (13.99%) were the main components which presented 67.91% of total

essential oil. Among the world, the 1,8-cineole chemotype characterises the *Rosmarinus officinalis* essential oil. Its concentration was ranged from 41 to 63% (Chalchat *et al.*, 1993; Fechtal *et al.*, 2001).

Chemical composition of the Portugal *Rosmarinus officinalis* essential oil is different of the Moroccan one collected in the same period (Faleiro *et al.*, 1999). This could be due to several factors particularly to the geographic region or environmental conditions (Cavaleino *et al.*, 2001; Angioni *et al.*, 2003).

The tested bacteria were all sensitive to *R. officinalis* essential oil in steam phase using the micro-atmospheric method. *P. vulgaris* was the most sensitive however *K. pneumonia* was the most resistant bacteria. It is interesting to note that *Rosmarinus officinalis* essential oil manifested important antibacterial activity against *E. coli* and *S. enteritidis*, which are known to be very resistant bacteria to synthetic drugs (Mimica *et al.*, 2003). Several components of the essential oils seem to contribute to the antimicrobial activity and there is no major component solely responsible for a such property (Faleiro *et al.*, 2003).

Prolonged treatment of mice allowed us to determine the functional and anatomopathological changes consecutive to the repeated administration of rosemary essential oil. The weight evolution and consumption of food and water showed absence of difference at the treated mice and witnesses during the treatment. We would conclude the absence of toxicity by Moroccan *Rosmarinus officinalis* essential oil during the treatment by oral administration. This in agrees with Lemonica *et al.* (1996).

The red color of lungs would indicate a better blood irrigation, probably by stimulation of the arteriolo-capillary system and that could be the cause of the alveolar dilation observed in lung. In fact, several studies showed that rosemary intervenes in relieving respiratory disorders and in prevention of asthma and cardio-vascular diseases (Nasel *et al.*, 1994; Al-Sereiti *et al.*, 1999; Aghel *et al.*, 2004). It is also a stimulant of suprarenal glands, what explain the hypertrophy of cells of suprarenal glands observed in our study. This histopathological investigations are the originality of the present paper.

We have conclude that *Rosmarinus officinalis* essential oil present no toxicity at 50 µl/g dose and possess stimulating effects of the suprarenal glands and of the respiratory system.

ACKNOWLEDGMENTS

We gratefully acknowledge helpful of Mme A. DADOUN from Bacteriological Laboratory, Centre of Biologic Medical, Pasteur Institute, Casablanca, Morocco and Mr M. CHEHEB, Laboratoire des Substances Naturelles et Arômes Alimentaires, Institut Agronomique et Vétérinaire Hassan II, Rabat.

REFERENCES

- Aghel N, Yamini Y, Hadjiakhondi A & Pourmortazavi SM (2004) Supercritical carbon dioxide extraction of *Mentha pulegium* L. essential oil. *Talanta* 62: 407-411
- Al-Sereiti M.R, Abu-amer K.M & Sen P (1999) Pharmacology of rosemary (*Rosmarinus officinalis*). *Indian J Exp Biol* 37 (2): 124-130
- Angioni A, Barra A, Russo MT, Coroneo V, Dessi S & Cabras P (2003) Chemical composition of the essential oils of juniperus from ripe and unripe berries and leaves and their antimicrobial activity. *J Agric Food Chem* 51 (10): 3073-3078
- Benkeblia N (2004) Antimicrobial activity of essential oil extracts of various onions (*Allium cepa*) and garlic (*Allium sativum*). *Lensm.-Wiss u.-Technol* 37: 263-268
- Calabrese V, Scapagnini G, Catalano C, Bate TE, Dinotta F, Micali G & Giuffrida Stella AM (2001) Induction of heat shock protein synthesis in human skin fibroblasts in response to oxidative stress: regulation by a natural antioxidant from rosemary extract. *Int J Tissue Reactv* XXIII (2): 51-58
- Cavaleino C, Rezzi S, Salgueiro L, Bighelli A, Casanova J & Da Cunha AP (2001) Intraspecific chemical variability of the leaf essential oil of *Juniperus phoenicea* var. *turbinata* from Portugal. *Biochem Syst Ecol* 29: 179-188
- Chalchat JC, Gary RP, Michet A, Benjilali B & Chabart P (1993) Essential oils of rosemary (*Rosmarinus officinalis* L.). The chemical composition of oils of various origins (Morocco, Spain, France). *Journal of essential oil research* 5 (6): 613-618
- Cosentino S, Tuberoso CIG, Pisano B, Satta M, Mascia V, Arzedi E & Palmas F (1999) *In vitro* antimicrobial activity and chemical composition of Sardinian Thymus essential oils. *Letters in Applied Microbiology* 29: 130-135
- Cowan MM (1999) Plant products as antimicrobial agents. *Clin Microbiol Rev* 12: 564-582
- Dorman HJ & Deans SG (2000) Antimicrobial agents from plants: antibacterial activity of plant volatile oils. *J Appl Microbiol* 88: 308-16
- Faleiro L, Miguel GM, Guerrero CAC & Brito JMC (1999) Antimicrobial activity of essential oils of *Rosmarinus officinalis* L., *Thymus mastichina* (L) L.SSP *mastichina* and *Thymus albicans* hofmanns e link Pharmacognosy. *Acta Horticulturae* 501: 445-448
- Faleiro M.L, Miguel M.G, Ladeiro F, Venâncio F, Tavares R, Brito JC, Fueiredo AC, Barroso JG & Pedro LG (2003) Antimicrobial activity of essential oils isolated from Portuguese endemic species of Thymus. *Letters in Applied Microbiology* 36: 35-40
- Fechtal M, Ismaili R & Zine El Adine A (2001) Effet de la transplantation sur la qualité et le rendement en huiles essentielles du romarin (*Rosmarinus officinalis*). *Ann Rech For (Maroc)* 34: 94-102
- Gijzen M, Efraim L, Savage T & Croteau R (1991) Conifer monoterpenes-biochemistry and bark beetle chemical ecology. In Bioactive Volatile compounds from Plants, ed. Teranishi R, Buttery RG & Sugisawa HCh, Washington DC: *American Chemistry Society*
- Kalembe D & Kunicka A (2003) Antibacterial and Antifungal Properties of Essential Oils. *Current Medicinal Chemistry* 10: 813-829
- Kim HK & Kim K (2003) Protein glycation inhibitory and antioxidative activities of some plant extracts *in vitro*. *J Agric Food Chem* 51 (6): 1586-1591
- Larrondo JV, Agut M & Calvo-Torras MA (1995) Antimicrobial activity of essences from labiates. *Microbios* 82: 171-172
- Lemonica IP, Damasceno DC & Di-stasi LC (1996) Study of the embryotoxic effects of an extract of rosmar (*Rosmarinus officinalis*). *Braz J Med Biol Res* 29 (2): 223-227
- Lis-Balchin M & Deans SG (1997) Bioactivity of selected plant essential oils against *Listeria monocytogenes*. *J Appl Microbiol* 82: 759-762
- Marino M, Bersani C & Comi G (2001) Impedance measurements to study the antimicrobial activity of essential oils from Lamiaceae and Compositae. *International Journal of Food Microbiology* 67: 187-195
- Mimica-Dukic N, Bozin B, Sokovic M, Mihajlovic B & Matavulj M (2003) Antimicrobial and antioxidant activities of three Mentha species essential oils. *Planta Med* 69: 413-419

- Mondello F, De Bernardis F, Girolamo A, Salvatore G & Cassone A (2003) *In vitro* and *in vivo* activity of tea tree oil against azole-susceptible and -resistant human pathogenic yeasts. *J Antimicrobial Chemotherapy* 51: 1223-1229
- Munne-Bosch S & Alegre L (2001) Subcellular compartmentation of the diterpene carosic acid and its derivatives on the leaves of rosemary. *Plant Physiol* 125 (2): 1094-1102
- Nasel C, Nasel B, Samec P, Schindler E & Buchbauer G (1994) Functional imaging of effects of fragrances on the human brain after prolonged inhalation. *Chem Senses* 19 (4): 359-364
- Offord EA, Mace K, Ruffieux C, Malnoe A & Pfeifer AM (1995) Rosemary components inhibit benzo[a]pyrene-induced genotoxicity in human bronchial cells. *Carcinogenesis* 16 (9): 2057-2062
- Pattanaik S, Subramanyam VR & Kole C (1996) Antibacterial and antifungal activity of the essential oils *in vitro*. *Microbios* 86 (349): 237-246
- Ponce AG, Del Valle CE & Roura SI (2004) Natural essential oils as reducing agents of peroxidase activity in leafy vegetables. *Lebensm- Wiss u - Technol* 37: 199-204
- Silva J, Abebe W, Sousa SM, Durte VG, Machado MIL & Matos FJA (2003) Analgesic and anti-inflammatory effects of essential oils of *Eucalyptus*. *J Ethnopharmacology* 89: 277-283
- Sotelo-Felix J.I, Martinez-Fong D, Muriel P, Santillan R.L, Castillo D & Yahuca P (2002) Evaluation of the effectiveness of *Rosmarinus officinalis* (Lamiaceae) in the alleviation of carbon tetrachloride-induced acute hepatotoxicity in the rat. *J Ethnopharmacol* 81 (2): 145-154
- Vadar-Unlu G, Candan F, Sokmen A, Daferera D, Polissiou M, Sokmen M, Donmez E & Tepe B (2003) Antimicrobial and antioxidant activity of the essential oil and methanol extracts of *Thymus pectinatus* Fisch. et Mey. Var. *pectinatus* (Lamiaceae). *J Agric Food Chem* 51: 63-67
- Zeng HH, Tu PF, Zhou K, Wang H, Wang BH & Lu JF (2001) Antioxidant properties of phenolic diterpenes from *Rosmarinus officinalis*. *Acta Pharmacol Sin* 22 (12): 1094-1098